

Elevated atmospheric CO₂ and soil N fertility effects on growth, mycorrhizal colonization, and xylem water potential of juvenile ponderosa pine in a field soil

R.F. Walker¹, D.R. Geisinger¹, D.W. Johnson² & J.T. Ball²

¹University of Nevada, Department of Environmental and Resource Sciences, Reno, NV 89512, USA*, and

²Desert Research Institute, Biological Sciences Center, Reno, NV 89506, USA

Received 17 September 1996. Accepted in revised form 15 May 1997

Key words: carbon dioxide, ectomycorrhizae, *Pinus ponderosa*, root systems, soil nitrogen, water relations

Abstract

Interactive effects of atmospheric CO₂ enrichment and soil N fertility on above- and below-ground development and water relations of juvenile ponderosa pine (*Pinus ponderosa* Dougl. ex Laws.) were examined. Open-top field chambers permitted creation of atmospheres with 700 $\mu\text{L L}^{-1}$, 525 $\mu\text{L L}^{-1}$, or ambient CO₂ concentrations. Seedlings were reared from seed in field soil with a total N concentration of approximately 900 $\mu\text{g g}^{-1}$ or in soil amended with sufficient (NH₄)₂SO₄ to increase total N by 100 $\mu\text{g g}^{-1}$ or 200 $\mu\text{g g}^{-1}$. The 525 $\mu\text{L L}^{-1}$ CO₂ treatment within the intermediate N treatment was excluded from the study. Following each of three consecutive growing seasons, whole seedlings of each combination of CO₂ and N treatment were harvested to permit assessment of shoot and root growth and ectomycorrhizal colonization. In the second and third growing seasons, drought cycles were imposed by withholding irrigation during which predawn and midday xylem water potential and soil water potential were measured. The first harvest revealed that shoot weight and coarse and fine root weights were increased by growth in elevated CO₂. Shoot and root volume and weights were increased by CO₂ enrichment at the second harvest, but growth stimulation by the 525 $\mu\text{L L}^{-1}$ CO₂ concentration exceeded that in 700 $\mu\text{L L}^{-1}$ CO₂ during the first two growing seasons. At the third harvest, above- and below-ground growth increases were largely confined to the 700 $\mu\text{L L}^{-1}$ CO₂ treatment, an effect accentuated by high soil N but evident in all N treatments. Ectomycorrhizal formation was reduced by elevated CO₂ after one growing season, but thereafter was not significantly affected by CO₂ and was unaffected by soil N throughout the study. Results of the xylem water potential measurements were variable, as water potentials in seedlings grown in elevated CO₂ were intermittently higher on some measurement days but lower on others than that of seedlings grown in the ambient atmosphere. These results suggest that elevated CO₂ exerts stimulatory effects on shoot and root growth of juvenile ponderosa pine under field conditions which are somewhat dependent on N availability, but that temporal variation may periodically result in a greater response to a moderate rise in atmospheric CO₂ than to a doubling of the current ambient concentration.

Introduction

Research to date concerned with the effects of atmospheric CO₂ enrichment on forest ecosystems has generally emphasized above-ground growth processes of tree seedlings, and the results have indicated that the stimulation of shoot growth by elevated CO₂ varies somewhat by species in both its magnitude and dura-

tion (Bazzaz et al., 1990; Brown and Higginbotham, 1986; Conroy et al., 1986; Kaushal et al., 1989; Norby and O'Neill, 1989; Norby et al., 1996; O'Neill et al., 1987a; Radoglou and Jarvis, 1990; Surano et al., 1986; Tissue et al., 1996). Due to the difficulty inherent in the examination of below-ground processes, less research has focused on effects of rising CO₂ on tree root systems. Although in some studies CO₂ enrichment promoted disproportionately greater root than shoot growth (Norby et al., 1986; O'Neill et

* FAX No: +17027844583. E-mail: Walker@ers.unr.edu

al., 1987a; Walker et al., 1995a, b), others revealed that stimulation of below-ground tree growth was a short-term response only (Brown and Higginbotham, 1986; Radoglou and Jarvis, 1990). For effects of elevated CO₂ on mycorrhizal colonization in forest tree seedlings, a positive response was observed by Norby et al. (1987), and positive but ephemeral responses by Lewis et al. (1994) and O'Neill et al. (1987b). As root systems provide critical feedback to the whole-tree response by altering the capacity to acquire water and nutrients (Norby, 1994), greater elucidation of the impact of rising CO₂ on root development is deemed crucial.

Comparatively little research has focused on the impact of elevated CO₂ on water relations of forest trees. Documented studies in this realm have consisted of pot studies which demonstrated that CO₂ enrichment may ameliorate the effects of water stress on seedling growth. Hollinger (1987) and Conroy et al. (1988) reported increases in instantaneous transpiration efficiency, while the results of related experiments (Conroy et al., 1986, 1988; Norby et al., 1986; Norby and O'Neill, 1989; Tolley and Strain, 1984) indicate improvement of whole-seedling water use efficiency in CO₂-enriched atmospheres.

The study reported here concerns the response of juvenile ponderosa pine (*P. ponderosa* Dougl. ex Laws.) to the interaction of elevated atmospheric CO₂ and soil N fertility under field conditions, with emphasis on root system development and water relations. We hypothesized that 1) CO₂ enrichment would stimulate above- and below-ground growth of ponderosa pine, an effect that would be accentuated by increased soil N availability, and 2) partitioning of this added growth to favor below-ground organs would enhance the resource acquisition capability of the root systems sufficiently to ameliorate the impact of short-term drought cycles on moisture stress.

Materials and methods

This study was conducted at the USDA Forest Service Institute of Forest Genetics (Placerville, CA) on a level uniform site at an elevation of 843 m with a soil of Aiken clay loam, a Xeric Haplohumult derived from andesite. Prior to the installation of the study, the site was occupied by a sparse grass cover, and the surrounding vegetation consists primarily of mature ponderosa pine trees. Site preparation consisted solely of disking to a depth of 25 cm. In 1991, 24 hexagonal open-top

chambers, each 3.6 m in diameter, were constructed of clear acrylic plastic sheeting to a height of 2.5 m with frustums of the same material extending above the chamber walls. These chambers were designed so that CO₂ could be pumped in from a central reservoir through a plenum at a set-point concentration. Selected concentrations were 700 $\mu\text{L L}^{-1}$ CO₂, 525 $\mu\text{L L}^{-1}$ CO₂, and ambient CO₂, with the latter predetermined as 352 $\mu\text{L L}^{-1}$. Automated monitoring of each chamber atmosphere revealed that actual CO₂ concentrations did not vary from the set points by more than 14 $\mu\text{L L}^{-1}$, and eight chambers were dedicated to each atmosphere. The distance between chambers was 5.5 m to prevent intermixing of the atmospheres. Irrigation water was delivered to each chamber via a standpipe to a looped 2.5-cm diameter manifold and distributed through low pressure spray heads on a schedule regulated by an electronic timer. The irrigation regime consisted of one 30-min period on each of three days per week from April through October, and approximately 0.7 cm of water was applied to each chamber during individual periods.

Intermediate and high soil N fertility treatments were imposed within individual chambers by annual application of sufficient (NH₄)₂SO₄ to increase total soil N by 100 $\mu\text{g g}^{-1}$ and 200 $\mu\text{g g}^{-1}$, respectively. The low N treatment consisted of unamended soil which had a total N concentration of approximately 900 $\mu\text{g g}^{-1}$. Total soil N was monitored periodically using a Perkin-Elmer 2400 CHN Analyzer (Perkin-Elmer Corp., Norwalk, CT) with samples collected by coring to a depth of 30 cm, and its concentration was consistent across the study site prior to the initial (NH₄)₂SO₄ applications. Three chambers were dedicated to each combination of CO₂ and N treatment, but due to the limitation imposed by the number of available chambers, the intermediate N regime within the 525 $\mu\text{L L}^{-1}$ CO₂ treatment was omitted from the study.

Half sib ponderosa pine seed (Eldorado County, CA seed source) were stratified at 3 °C for 30 days and then sterilized in 10% H₂O₂ for 10 min. In May 1991, 66 pretreated seeds were sown per chamber in groups of three on a 30-cm square spacing. These seeds were sown directly into the soil constituting the chamber floors so that root egress would not be constrained by artificial barriers. Irrigation was applied as needed to induce germination, which was followed by a thinning to 22 equally-spaced seedlings per chamber.

Following the 1991 and 1992 growing seasons, three whole seedlings from each chamber were har-

vested per year using water pressure to excavate complete root systems. One additional seedling from each chamber was harvested by this method following the 1993 growing season. Thus, the first, second, and third harvests were completed 6, 18, and 30 months, respectively, after the chambers were seeded. At each harvest, individual roots of the selected seedlings were excavated from the root collar to their terminus to provide assurance that complete root systems were obtained. This frequently necessitated sacrificing seedlings immediately adjacent to those selected for harvest, which increased the spacing between the remaining seedlings, but equal numbers of seedlings were maintained in each chamber throughout the study and the seedlings within individual chambers remained approximately equidistant. Immediately after each harvest, shoots were severed at the root collar, the root systems were washed, and shoot and root volumes were determined by water displacement using the apparatus of Yawney and Carl (1969).

Ectomycorrhizal colonization of harvested root systems was quantified by measuring the total length of each root system and counting the root segments (1.0 cm lengths) exhibiting characteristic monopodial, bifurcate, or coralloid short roots, or short roots with an obvious fungal mantle. The number of root segments with mycorrhizae was then expressed as a percentage of the total length of the root system (Grand and Harvey, 1982). Ectomycorrhizae resulting from infection by different mycobionts were grouped together in the quantification procedure, but the fungi forming these mycorrhizae were identified as to probable species when possible by the sporocarps appearing in the chambers and by mycorrhizal color and morphology. Following mycorrhizal quantification, the shoots and root systems of each seedling were dried at 75 °C for 36 hours and weighed. Root systems were divided into coarse (≥ 2.0 mm diameter) and fine (< 2.0 mm diameter) root fractions which were weighed and recorded separately but then combined for calculation of shoot/root ratios, whole-seedling volume/weight ratios, and specific root length.

In September 1992, predawn and midday measurements of xylem water potential (Ψ_p) were made on seedlings of all treatments by the method of Waring and Cleary (1967) using a PMS Model 600 portable pressure chamber (PMS Instrument Co., Corvallis, OR). These measurements were made on the first, third, fifth, and seventh day after discontinuing irrigation in order to examine seedling water relations as affected by treatment during short-term imposed drought episodes.

Predawn measurements were completed between 0500 and 0600 hours and midday measurements were completed between 1300 and 1400 hours. During each measurement period, a needle from the upper crown of one seedling from every chamber was selected for measurement. Upon excision, the needles were immediately placed in the chamber, and the chamber pressure was then increased at a constant rate of 0.03 MPa s^{-1} until sap appeared uniformly over the cut surface. Between the predawn and midday Ψ_p measurements of each day, soil water potential (Ψ_s) was measured with a Soilmoisture Model 5910-A digital soil moisture meter and Model 5201 gypsum blocks (Soilmoisture Equipment Corp., Santa Barbara, CA). One gypsum block had been previously installed to a depth of 20 cm at the center of each chamber. The series of Ψ_p and Ψ_s measurements described above was repeated in May, July, and September of 1993 in order to examine seasonal differences in the effect of the CO_2 and N treatments on seedling water relations.

Two-way analysis of variance was performed on all data derived from this 3×3 (three atmospheric CO_2 \times three soil N treatments) incomplete factorial experiment, with the intermediate soil N and $525 \mu\text{L L}^{-1}$ CO_2 treatment combination excluded. Type IV sum of squares was used to accommodate the missing treatment combination, and the CO_2 and N main treatment and $\text{CO}_2 \times \text{N}$ interaction effects were considered significant when $p \leq 0.05$ according to the F test. The arcsine transformation was performed on the mycorrhizal infection percentages prior to analysis. Data derived from each of the three harvests and each of the four series of water relations measurements were analyzed separately, with the data collected from seedlings in individual chambers constituting replications and thus three replications per treatment combination. All statistical analyses were accomplished using the Statistical Analysis System (SAS Institute, Inc., Cary, NC). In the presentation of results that follows, p values are included in the text when either a main treatment or treatment interaction effect proved significant as determined through analysis of variance. Also, the standard error (SE) of the mean for all variables presented in tables is included to provide an indication of the variation within treatment means, and the standard error of the difference (SED) among means for both main treatment and treatment interaction effects is included to facilitate comparisons among means for all growth variables.

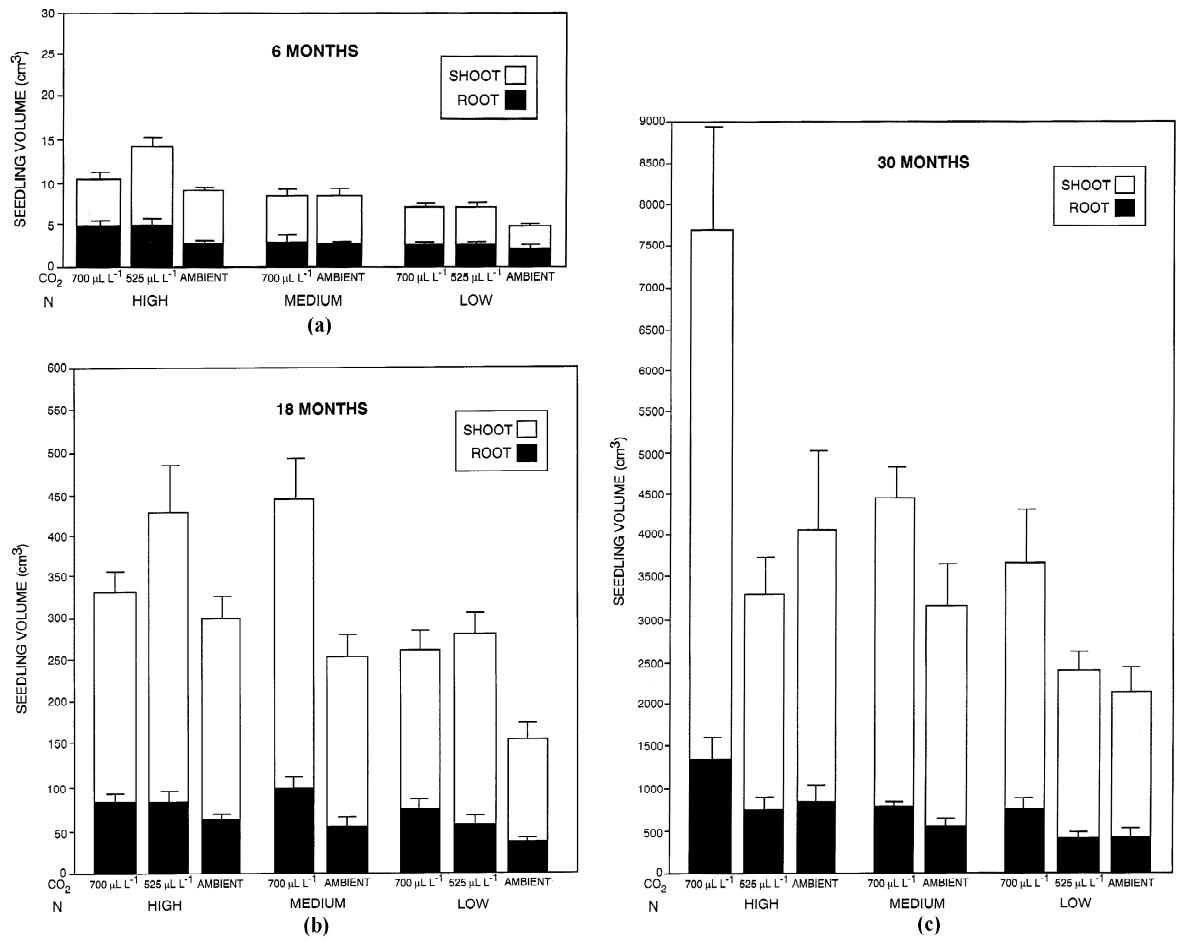


Figure 1. Shoot and root volumes after 6, 18, and 30 months of juvenile ponderosa pine as affected by atmospheric CO₂ and soil N concentration. Bars represent the SE of each mean. For shoot volume at 6 months, SED = 1.5 for CO₂, 1.5 for N, and 2.5 for CO₂ × N; for root volume at 6 months, SED = 0.8 for CO₂, 0.8 for N, and 1.3 for CO₂ × N; for shoot volume at 18 months, SED = 58 for CO₂, 58 for N, and 94 for CO₂ × N; for root volume at 18 months, SED = 15 for CO₂, 15 for N, and 25 for CO₂ × N; for shoot volume at 30 months, SED = 876 for CO₂, 876 for N, and 1430 for CO₂ × N; and for root volume at 30 months, SED = 123 for CO₂, 123 for N, and 201 for CO₂ × N.

Results

Shoot growth

Analysis of variance revealed that shoot volume was significantly increased ($p = 0.0245$) by N fertilization but was unaffected by elevated CO₂ after one growing season (Figure 1). After 18 months, however, both the CO₂ ($p = 0.0298$) and N ($p = 0.0129$) treatments significantly influenced shoot growth, as a substantial volume increase was evident in the 525 μL L⁻¹ CO₂ treatment in high soil N, in the 700 μL L⁻¹ CO₂ treatment in medium soil N, and in both enriched atmospheres in low N. Furthermore, the smallest shoot volumes after two growing seasons were those of seedlings grown in

low soil N and ambient CO₂. After 30 months, shoot volume was significantly increased ($p = 0.0411$) by the high CO₂ atmosphere in all three N treatments. The greatest response was evident in high soil N, however, as the volume of these seedlings exceeded that of any other treatment by $\geq 73\%$. Unlike the high CO₂ concentration, the 525 μL L⁻¹ CO₂ atmosphere did not increase shoot volume after three growing seasons in high soil N and produced only a marginal increase in low N.

Although shoot volume was not significantly affected by the CO₂ concentration after 6 months, shoot dry weight was increased ($p = 0.0498$) by both the 700 μL L⁻¹ and 525 μL L⁻¹ CO₂ atmospheres, with the latter having a greater stimulatory effect than the for-

mer in high and low N (Table 1). Shoot weight was also affected by N fertility after one growing season ($p = 0.0027$), with the high N treatment producing the greatest biomass. The above pattern of treatment effects persisted through 18 months, as both CO₂ enrichment ($p = 0.0007$) and N fertilization ($p = 0.0187$) increased shoot weight, and seedlings grown in the intermediate CO₂ concentration were again larger than those grown in high CO₂ in high and low N. Unlike the first growing season, however, high CO₂ substantially increased shoot weight in the medium N treatment as well. At the third harvest, a shift in the effect of atmospheric CO₂ was revealed, as increases in shoot weight were largely a response ($p = 0.0417$) to the 700 $\mu\text{L L}^{-1}$ concentration alone. Shoot weight was also increased by N fertility ($p = 0.0296$) after 30 months, and overall, the combination of high N and high CO₂ produced the greatest shoot biomass by a substantial margin.

Root growth

Root volume was not significantly affected by either atmospheric CO₂ or N fertility after 6 months (Figure 1). During the second growing season, however, elevated CO₂ ($p = 0.0202$) and N fertilization ($p = 0.0499$) stimulated root volume growth, as a stepwise response to increasing CO₂ was evident in medium and low N after 18 months while the 700 $\mu\text{L L}^{-1}$ and 525 $\mu\text{L L}^{-1}$ CO₂ atmospheres produced a comparable growth increase in high N. Low soil N produced the smallest root volumes overall after the second growing season, although the ambient atmosphere resulted in root growth in high and medium soil N similar to that in low N. The third harvest revealed a root growth response to CO₂ enrichment parallel to that of the shoots, as the high CO₂ atmosphere increased root volume ($p = 0.0077$) but this effect did not extend to the medium CO₂ concentration. Soil N also had a significant effect ($p = 0.0097$) on root growth, as root volumes generally increased with increasing N availability. Overall, the largest root volume was that of seedlings grown in high soil N and the 700 $\mu\text{L L}^{-1}$ CO₂ atmosphere, which exceeded that of any other treatment by $\geq 56\%$ after 30 months.

Elevated CO₂ and N fertilization increased root dry weight after one growing season in both the coarse ($p = 0.0464$ for CO₂ treatment and $p = 0.0015$ for N treatment) and fine ($p = 0.0414$ and $p = 0.0216$ for CO₂ and N treatments, respectively) root fractions (Table 1). The smallest root systems within N treatments were those of seedlings grown in ambient CO₂ and

the smallest of any treatment were those of seedlings grown in ambient CO₂ and low N for both root fractions. The second harvest revealed a similar response in coarse root growth to the CO₂ ($p = 0.0026$) and N ($p = 0.0146$) concentrations. In the fine root fraction, however, CO₂ enrichment alone resulted in an increase ($p = 0.0270$) in dry weight, and the 525 $\mu\text{L L}^{-1}$ CO₂ atmosphere did not stimulate fine root growth in low soil N. Root weight measurements after 30 months indicated a modification of treatment effects, although here also the CO₂ ($p = 0.0219$) and N ($p = 0.0128$) concentration effects were significant for coarse root growth. However, the increase in coarse root weight was largely confined to the high CO₂ treatment regardless of N availability, and neither CO₂ nor N significantly affected fine root growth. Nevertheless, the overall result of the root weight measurements after three growing seasons indicated a response in concert with that revealed previously for above- and below-ground growth, as the largest combined coarse and fine root weight of any treatment was that of seedlings grown in high CO₂ and high N.

Examination of specific root length revealed that the effect of CO₂ concentration on this root growth variable was nonsignificant throughout the study (Table 1). Specific root length was also unaffected by N treatment after 6 months, but was increased by low soil N after 18 months ($p = 0.0339$) and by medium N after 30 months ($p = 0.0133$).

Shoot/root and volume/weight ratios

Shoot/root ratio was not significantly affected by either CO₂ enrichment or N fertilization at the first and third harvests (Table 1). The second harvest revealed a conflicting, although significant ($p = 0.0454$), result with regard to elevated CO₂, however, as the lowest shoot/root ratios within the high and low N treatments were those of seedlings grown in 700 $\mu\text{L L}^{-1}$ CO₂, but this atmosphere produced the highest ratio in medium N. Similarly, whole-seedling volume/weight ratio was unaffected by CO₂ and N after the first and third growing seasons, but volume/weight ratios were significantly higher ($p = 0.0002$) in the ambient CO₂ treatment after 18 months.

Mycorrhizal colonization

The initial harvest revealed that the percentage of total root system length colonized by ectomycorrhizal fungi was reduced by elevated CO₂ ($p = 0.0471$), as the

Table 1. Dry weight and ectomycorrhizal colonization after 6, 18, and 30 months of juvenile ponderosa pine as affected by atmospheric CO₂ and soil N concentration. The SE of each mean is in parentheses

Harvest	Soil N and CO ₂ treatment	Shoot weight (g)	Root weight (g)		Shoot/root ratio	Volume/weight ratio (cm ³ g ⁻¹)	Specific root length (m g ⁻¹)	Mycorrhizal infection (%)	
			Coarse	Fine					
6 months	<i>High soil N</i>								
	700 μL L ⁻¹ CO ₂	1.57 (0.26)	0.32 (0.05)	0.37(0.06)	2.25(0.21)	5.06(0.37)	3.09(0.34)	22(3)	
	525 μL L ⁻¹ CO ₂	1.84 (0.33)	0.31 (0.07)	0.37(0.05)	2.68(0.25)	5.79(0.64)	3.83(0.48)	20(3)	
	Ambient CO ₂	1.28 (0.17)	0.20 (0.03)	0.26(0.04)	2.91(0.27)	7.53(2.52)	4.51(0.38)	32(9)	
	<i>Medium soil N</i>								
	700 μL L ⁻¹ CO ₂	0.95 (0.24)	0.17 (0.06)	0.28(0.05)	2.13(0.19)	5.33(0.63)	4.68(1.38)	24(3)	
	Ambient CO ₂	0.94 (0.20)	0.11 (0.02)	0.23(0.03)	2.74(0.29)	7.36(0.74)	4.78(0.20)	50(8)	
	<i>Low soil N</i>								
	700 μL L ⁻¹ CO ₂	0.81 (0.14)	0.14 (0.03)	0.22(0.04)	2.26(0.24)	6.83(0.63)	3.87(0.29)	27(4)	
	525 μL L ⁻¹ CO ₂	1.03 (0.17)	0.16 (0.03)	0.26(0.02)	2.44(0.24)	5.11(0.58)	4.09(0.47)	45(7)	
	Ambient CO ₂	0.54 (0.07)	0.08 (0.01)	0.17(0.01)	2.09(0.15)	6.77(0.77)	4.84(0.60)	52(9)	
	SED: CO ₂	0.32	0.06	0.06	0.35	1.61	0.94	10	
	N	0.32	0.06	0.06	0.35	1.61	0.94	10	
	CO ₂ × N	0.52	0.10	0.10	0.58	2.63	1.53	16	
	18 months	<i>High soil N</i>							
700 μL L ⁻¹ CO ₂		75.43 (10.29)	23.24 (3.04)	4.08(0.47)	2.82(0.26)	3.39(0.15)	0.59(0.08)	27(6)	
525 μL L ⁻¹ CO ₂		107.82 (18.56)	27.93 (4.42)	4.09(0.85)	3.41(0.40)	3.20(0.14)	0.46(0.04)	35(3)	
Ambient CO ₂		56.27 (7.34)	16.14 (1.95)	2.37(0.16)	3.01(0.16)	4.09(0.07)	0.56(0.07)	29(4)	
<i>Medium soil N</i>									
700 μL L ⁻¹ CO ₂		94.79 (15.95)	29.16 (5.50)	3.28(0.25)	3.00(0.21)	3.57(0.09)	0.39(0.04)	29(6)	
Ambient CO ₂		47.75 (8.96)	13.67 (2.31)	3.23(0.39)	2.69(0.18)	4.12(0.16)	0.70(0.11)	30(4)	
<i>Low soil N</i>									
700 μL L ⁻¹ CO ₂		53.12 (11.83)	17.35 (3.62)	3.14(0.45)	2.57(0.27)	3.92(0.25)	0.65(0.09)	30(6)	
525 μL L ⁻¹ CO ₂		68.28 (9.02)	17.58 (2.70)	2.45(0.33)	4.05(0.94)	3.25(0.24)	0.53(0.09)	30(6)	
Ambient CO ₂		26.90 (3.28)	6.76 (1.09)	2.50(0.29)	3.18(0.56)	4.65(0.60)	0.91(0.18)	30(4)	
SED: CO ₂		17.43	5.04	0.67	0.67	0.39	0.15	8	
N		17.43	5.04	0.67	0.67	0.39	0.15	8	
CO ₂ × N		28.46	8.22	1.10	1.09	0.64	0.24	12	
30 months		<i>High soil N</i>							
	700 μL L ⁻¹ CO ₂	2118.9 (696.1)	378.2 (82.1)	16.6(5.6)	5.1(0.8)	3.0 (0.1)	0.2 (0.1)	41 (8)	
	525 μL L ⁻¹ CO ₂	1036.5 (95.2)	204.0 (44.9)	18.5(3.1)	5.0(0.8)	2.6 (0.2)	0.2 (0.1)	27(3)	
	Ambient CO ₂	1235.8 (260.4)	234.5 (55.0)	11.2(5.0)	5.1(0.3)	2.7 (0.3)	0.2 (0.1)	21(3)	
	<i>Medium soil N</i>								
	700 μL L ⁻¹ CO ₂	1294.8 (165.8)	223.3 (14.5)	22.0(2.4)	5.3(0.7)	2.9 (0.1)	0.3 (0.1)	34(15)	
	Ambient CO ₂	841.9 (167.0)	160.7 (23.3)	14.1(3.3)	4.7(0.3)	3.1 (0.1)	0.3 (0.1)	32(10)	
	<i>Low soil N</i>								
	700 μL L ⁻¹ CO ₂	1068.3 (183.8)	208.0 (37.4)	12.0(2.2)	4.9(0.1)	2.9 (0.3)	0.2 (0.1)	38(8)	
	525 μL L ⁻¹ CO ₂	685.3 (93.3)	135.3 (32.2)	9.1(2.0)	5.0(0.6)	2.9 (0.1)	0.2 (0.1)	35(7)	
	Ambient CO ₂	619.0 (118.9)	123.5 (21.0)	10.9(3.3)	4.7(0.6)	2.9 (0.1)	0.2 (0.1)	49(11)	
	SED: CO ₂	251.3	38.0	3.1	0.5	0.2	0.1	8	
	N	251.3	38.0	3.1	0.5	0.2	0.1	8	
	CO ₂ × N	410.3	62.1	5.1	0.8	0.3	0.1	13	

highest infection percentages within each N treatment were those of seedlings grown in ambient CO₂ (Table 1). After 18 months, however, infection percentages were generally consistent across all treatments. The third harvest provided conflicting results regarding the influence of CO₂ on mycorrhizal colonization, as the highest infection percentage in the high N treatment was exhibited by seedlings grown in 700 μL L⁻¹ CO₂, while in low N, seedlings grown in the ambient atmosphere exhibited the highest infection level. Treatment effects were statistically nonsignificant after 30 months as well, however. The predominant mycobiont colonizing the roots of all seedlings throughout the study was *Suillus granulatus* (L. ex Fr.) Kuntze, readily identified due to the prevalence of its characteristic sporocarps (Lincoff, 1981) in the chambers and the color and morphology of its mycorrhizae (Riffle, 1973). Much less frequently encountered were ectomycorrhizae formed by *Cenococcum geophilum* Fr., which were identified by their unique coloration (Molina and Trappe, 1982).

Water relations

Substantial variability in predawn Ψ_p was evident both within and between the four series of water relations measurements (Figure 2). Predawn Ψ_p in the early fall of the second growing season was not affected on any of the four days of measurement by either the CO₂ or N treatment. Likewise, predawn measurements on the first and third days after discontinuing irrigation in the spring of the third growing season were unaffected by either treatment, but on the fifth day predawn Ψ_p of seedlings grown in 700 μL L⁻¹ CO₂ and of seedlings grown in high soil N and 525 μL L⁻¹ CO₂ were higher ($p = 0.0286$) than those of the other treatments. Also, predawn Ψ_p of unfertilized seedlings was lower, with the exception of those grown in 700 μL L⁻¹ CO₂, than that of fertilized seedlings on this measurement day ($p = 0.0329$). The series of Ψ_p measurements completed in the summer of the third growing season also revealed that predawn Ψ_p of seedlings grown in elevated CO₂ exceeded that of seedlings grown in the ambient atmosphere on the fifth day after cessation of irrigation ($p = 0.0129$). However, measurements completed in the early fall of the third growing season were consistent with those made in the fall of the previous year in that predawn Ψ_p was unaffected by either CO₂ or N treatment on all measurement days.

The intermittent and inconsistent pattern of statistically significant treatment effects observed in the predawn measurements was also noted in midday Ψ_p

(Figure 2). One day after suspending irrigation for the measurements made in the fall of the second growing season, midday Ψ_p of seedlings grown in 700 μL L⁻¹ CO₂ and that of the 525 μL L⁻¹ CO₂ treatment grown in high N were lower ($p = 0.0498$) than those of the other treatments, but this result did not persist through the remainder of this measurement series. Likewise, midday Ψ_p in the spring of the third growing season was not affected by CO₂ or N on any day of measurement. However, summer measurements of the third growing season revealed that midday Ψ_p of unfertilized seedlings grown in ambient CO₂ exceeded that of all other treatments on the seventh day after discontinuing irrigation ($p = 0.0476$ and $p = 0.0435$ for CO₂ and N treatments, respectively). Midday measurements made in the early fall of the third growing season revealed significant treatment effects on the third and seventh day after suspending irrigation. On the third day, midday Ψ_p of seedlings grown in elevated CO₂ was higher ($p = 0.0444$) than that of seedlings grown in the ambient atmosphere regardless of N treatment. Conversely, measurements on the seventh day revealed that midday Ψ_p of the elevated CO₂ treatments were lower ($p = 0.0432$) than that of the ambient treatment.

The Ψ_s measurements made in conjunction with the measurement of Ψ_p revealed that soil water availability did not differ significantly among the CO₂ or N treatments on any day within any series of the water relations measurements. Overall, Ψ_s declined on successive days of measurement after discontinuing irrigation within each of the four series (Table 2). Comparatively, higher Ψ_s values were encountered in the fall of the second growing season and lower values were prevalent in the spring of the third year, with an intermediate range of Ψ_s evident in the summer and fall of the third season.

Discussion

An array of results have been reported concerning the effect of CO₂ enrichment on shoot growth of forest tree seedlings. In previous experiments with ponderosa pine, which have primarily consisted of pot studies, Surano et al. (1986) observed an enhanced above-ground growth after 2.5 years in an atmosphere containing approximately twice the ambient CO₂ concentration. However, Walker et al. (1995a, b) found that 1.5×ambient CO₂ stimulated shoot growth of ponderosa pine seedlings more than a twice ambient atmosphere after one year if mineral nutrition was ade-

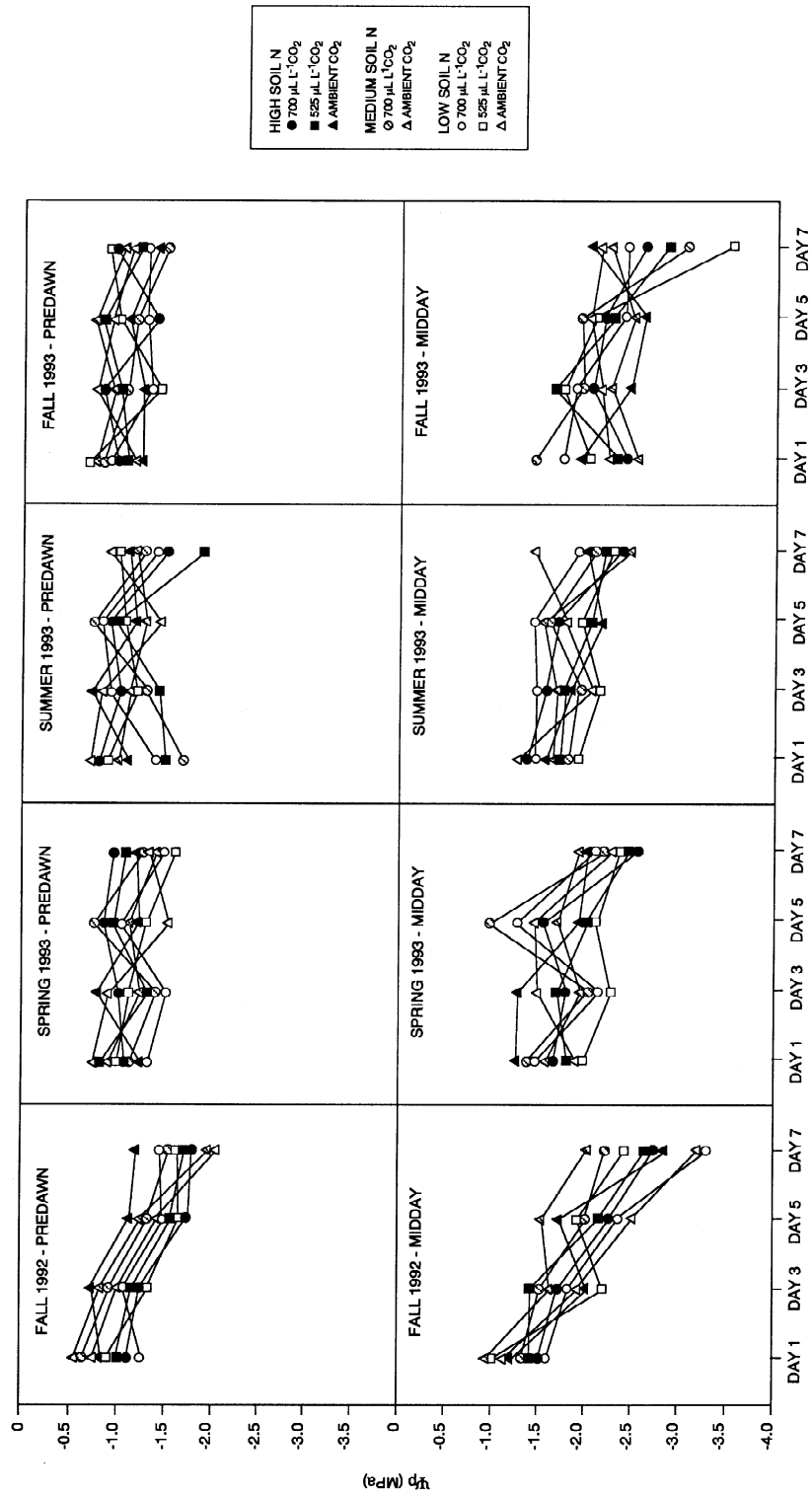


Figure 2. Predawn and midday xylem water potential (Ψ_p) of juvenile ponderosa pine as affected by atmospheric CO_2 and soil N concentration 1, 3, 5, and 7 days after cessation of irrigation in the fall of the second growing season and in the spring, summer, and fall of the third growing season.

Table 2. Soil water potential (Ψ_s) during measurements of xylem water potential in juvenile ponderosa pine as affected by atmospheric CO₂ and soil N concentration. The SE of each mean is in parentheses

Season and year of measurement	Soil N and CO ₂ treatment	Ψ_s (MPa)			
		Day 1	Day 3	Day 5	Day 7
Fall 1992	<i>High soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.002(0.001)	-0.008(0.002)	-0.006(0.001)	-0.009(0.001)
	525 $\mu\text{L L}^{-1}$ CO ₂	-0.002(0.001)	-0.011(0.007)	-0.007(0.002)	-0.014(0.007)
	Ambient CO ₂	-0.001 (0.001)	-0.006(0.001)	-0.004(0.001)	-0.007(0.002)
	<i>Medium soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.002(0.001)	-0.007(0.001)	-0.011(0.006)	-0.008(0.001)
	Ambient CO ₂	-0.001(0.001)	-0.005(0.001)	-0.005(0.001)	-0.010(0.001)
	<i>Low soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.001(0.001)	-0.008(0.003)	-0.006(0.001)	-0.007(0.002)
525 $\mu\text{L L}^{-1}$ CO ₂	-0.002(0.001)	-0.007(0.003)	-0.003(0.001)	-0.007(0.001)	
Ambient CO ₂	-0.001(0.001)	-0.005(0.001)	-0.003(0.002)	-0.008(0.002)	
Spring 1993	<i>High soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.082(0.047)	-0.053(0.019)	-0.137(0.035)	-1.167(0.167)
	525 $\mu\text{L L}^{-1}$ CO ₂	-0.067(0.032)	-0.052(0.012)	-0.452(0.216)	-1.052(0.448)
	Ambient CO ₂	-0.042(0.007)	-0.045(0.005)	-0.467(0.033)	-1.133(0.367)
	<i>Medium soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.038(0.008)	-0.040(0.006)	-0.102(0.049)	-0.685(0.414)
	Ambient CO ₂	-0.043(0.009)	-0.040(0.001)	-0.212(0.100)	-1.267(0.233)
	<i>Low soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.040(0.001)	-0.040(0.001)	-0.055(0.003)	-0.525(0.246)
525 $\mu\text{L L}^{-1}$ CO ₂	-0.123(0.088)	-0.082(0.037)	-0.160(0.0721)	-0.933(0.067)	
Ambient CO ₂	-0.033(0.003)	-0.033(0.003)	-0.163(0.118)	-0.510(0.257)	
Summer 1993	<i>High soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.047(0.017)	-0.050(0.020)	-0.057(0.022)	-0.098(0.033)
	525 $\mu\text{L L}^{-1}$ CO ₂	-0.033(0.003)	-0.033(0.003)	-0.153(0.123)	-0.062(0.020)
	Ambient CO ₂	-0.030(0.001)	-0.030(0.001)	-0.050(0.001)	-0.057(0.018)
	<i>Medium soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.027(0.003)	-0.027(0.003)	-0.030(0.006)	-0.090(0.055)
	Ambient CO ₂	-0.030(0.001)	-0.030(0.001)	-0.033(0.003)	-0.050(0.020)
	<i>Low soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.033(0.003)	-0.033(0.003)	-0.040(0.001)	-0.093(0.031)
525 $\mu\text{L L}^{-1}$ CO ₂	-0.027(0.003)	-0.027(0.003)	-0.040(0.006)	-0.147(0.053)	
Ambient CO ₂	-0.030(0.001)	-0.030(0.001)	-0.030(0.001)	-0.043(0.007)	
Fall 1993	<i>High soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.082(0.047)	-0.082(0.047)	-0.082(0.047)	-0.093(0.053)
	525 $\mu\text{L L}^{-1}$ CO ₂	-0.040(0.006)	-0.040(0.006)	-0.038(0.008)	-0.120(0.090)
	Ambient CO ₂	-0.030(0.001)	-0.033(0.003)	-0.033(0.003)	-0.033(0.003)
	<i>Medium soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.037(0.007)	-0.033(0.003)	-0.033(0.003)	-0.037(0.003)
	Ambient CO ₂	-0.030(0.001)	-0.033(0.003)	-0.033(0.003)	-0.033(0.003)
	<i>Low soil N</i>				
	700 $\mu\text{L L}^{-1}$ CO ₂	-0.040(0.001)	-0.040(0.001)	-0.037(0.003)	-0.040(0.001)
525 $\mu\text{L L}^{-1}$ CO ₂	-0.030(0.006)	-0.033(0.003)	-0.037(0.007)	-0.085(0.045)	
Ambient CO ₂	-0.037(0.007)	-0.037(0.007)	-0.037(0.007)	-0.040(0.006)	

quate. Results reported here also provide evidence of a greater shoot response in juvenile ponderosa pine to the 1.5× ambient than the 2× ambient atmosphere for the first two growing seasons. Predictions of plant reactions to increasing CO₂ derived from the photosynthesis model of Farquhar et al. (1980) indicate that growth may be temporarily restricted in high CO₂ by dilution of foliar N to the extent that photosynthetic capacity declines, a limitation not imposed by a moderate rise in CO₂ above the ambient concentration (Y Luo, Biological Sciences Center, Desert Research Institute, Reno, NV, personal communication). This may partially explain the higher growth rate of the 525 μL L⁻¹ CO₂ treatment relative to that in 700 μL L⁻¹ CO₂ during the first two growing seasons observed here. Nevertheless, increases in shoot growth were largely confined to the high CO₂ treatment after 30 months, a response common to all three N fertility treatments but accentuated by high N. This suggests that low soil N does not preclude a positive response to elevated CO₂ in ponderosa pine, but that the effect is magnified when N is readily available.

Most studies concerned with effects of CO₂ enrichment on the root systems of forest tree seedlings have revealed that below-ground growth was stimulated by elevated CO₂ (Rogers et al., 1994). Root volume and dry weight measurements presented here support this conclusion for ponderosa pine, although significant CO₂ treatment effects on volume did not appear until the second growing season and CO₂ effects on fine root weight did not persist through the final harvest. It has been hypothesized that CO₂ enrichment may accelerate growth of below-ground tissues proportionally more than those above ground in forest trees (Norby, 1989), and subsequent research has shown that this is true for some species (Rogers et al., 1994). However, there was little evidence in the data presented here that elevated CO₂ favors root growth over that of the shoots in ponderosa pine, as shoot/root ratios were largely unaffected by CO₂ treatment throughout the study. Furthermore, the accentuation of root growth in the 700 μL L⁻¹ CO₂ atmosphere by the high N treatment was less pronounced than that of the shoots after three growing seasons.

Ectomycorrhizal quantification of complete root systems harvested after 6 months of seedling growth revealed that the percentage of root length colonized decreased in elevated CO₂ relative to that of seedlings grown in the ambient atmosphere. Thereafter, atmospheric CO₂ was of little or no consequence in ectomycorrhizal formation, as all treatment effects were

nonsignificant after 18 and 30 months. Thus, results presented here are not in agreement with those of Norby et al. (1987) which indicated that ectomycorrhizal development of *P. echinata* Mill. was stimulated by a doubling of the ambient CO₂ concentration. Furthermore, they are not in concurrence with those of studies in which CO₂ enrichment resulted in a temporary increase in ectomycorrhizal colonization of *P. echinata* Mill., *Quercus alba* L. (O'Neill et al., 1987b), and *P. taeda* L. (Lewis et al., 1994) that did not persist through the end of the experiments. Previous research with ponderosa pine of somewhat longer duration than the above studies provided mixed results, including a finding of no CO₂ effect on ectomycorrhizal formation in one experiment (Walker et al., 1995a) but a significant increase in colonization after one year resulting from a doubling of ambient CO₂ in another (Walker et al., 1995b). It is apparent that the conclusions drawn from studies concerned with effects of CO₂ enrichment on the mycorrhizal development of forest trees is at least partially dependent upon the experimental conditions imposed as well as the species investigated. Given the episodic nature of ectomycorrhizal formation in most species, the duration of the experiment and the frequency and timing of sampling are particularly important in the interpretation of results. Nevertheless, the only reasonable conclusion that can be drawn from the data presented here is that CO₂ enrichment did not elevate ectomycorrhizal colonization of ponderosa pine during the first three years of growth.

The water relations measurements in the study reported here involved examination of the effects of growth in enriched CO₂ atmospheres on water stress of ponderosa pine in the field during drought episodes imposed in the early, middle, and late phases of the growing season. Overall, these results must be considered inconclusive, however, due to the intermittent pattern of significant treatment effects observed throughout the four series of predawn and midday Ψ_p measurements, and the propensity of seedlings grown in elevated CO₂ to exhibit a Ψ_p that was higher or lower than that of seedlings grown in the ambient atmosphere depending on when it was measured. This was exemplified in the last series of measurements, which were made at the end of the third growing season, when midday Ψ_p was higher in seedlings grown in elevated CO₂ on the third day after discontinuing irrigation but below that of the ambient treatment on the seventh day of this series. Furthermore, little can be deduced from these data about the linkage between proportionality of shoot and root biomass as affected by CO₂ treat-

ment and the response of ponderosa pine to drought, as CO₂ enrichment did not definitively alter shoot/root ratio in this study. It is possible that greater clarity of results may have been achieved if the duration of the drought episodes had been extended in order to impart a higher degree of water stress, although seedling Ψ_p measured here was generally well below that at which water potential-mediated plant processes are impacted (Hsiao et al., 1976).

Conflicting results have been presented concerning the potential fertilization effect of rising atmospheric CO₂ in mature natural forests (Graumlich, 1991; Kienast and Luxmoore, 1988; LaMarche et al., 1984). Likewise, the growing body of information derived from the studies with seedlings previously cited indicates substantial interspecific, intraspecific, and temporal variation in the response to CO₂ enrichment. Continued research on the impact of elevated CO₂ during all phases of tree growth is thus necessary before definitive conclusions regarding this phenomenon can be reached. The results presented here suggest that atmospheric CO₂ enrichment administered under field conditions stimulates proportional above- and below-ground growth of juvenile ponderosa pine, and the magnitude of this response is dependent on soil N availability. However, they also suggest that temporal variation in this response may periodically result in greater growth stimulation by a moderate CO₂ increase than by a doubling of the ambient concentration.

Acknowledgements

The authors thank L F Pitelka, P D Ross, A A Elwinger, D L Adkins, K W Sorom, W H Wright, D L Anderson, V H Yturiaga, and C A Johnson for their assistance. Partial support for this research was provided by the Electric Power Research Institute, Palo Alto, CA, to the Desert Research Institute and subsequently to the Nevada Agricultural Experiment Station Project 5256; additional support was provided by the Nevada Agricultural Experiment Station Projects 5242 and 522V funded by the McIntire-Stennis Cooperative Forestry Research Program.

References

Bazzaz F A, Coleman J S and Morse S R 1990 Growth responses of seven major co-occurring tree species of the northeastern United States to elevated CO₂. *Can. J. For. Res.* 20, 1479–1484.

- Brown K and Higginbotham K O 1986 Effects of carbon dioxide enrichment and nitrogen supply on growth of boreal tree seedlings. *Tree Physiol.* 2, 223–232.
- Conroy J, Barlow E W R and Bevege D I 1986 Response of *Pinus radiata* seedlings to carbon dioxide enrichment at different levels of water and phosphorus: Growth, morphology and anatomy. *Ann. Bot.* 57, 165–177.
- Conroy J P, Koppers M, Koppers B, Virgona J and Barlow E W R 1988 The influence of CO₂ enrichment, phosphorus deficiency and water stress on the growth, conductance and water use of *Pinus radiata* D. Don. *Plant Cell Environ.* 11, 91–98.
- Farquhar G D, von Caemmerer S and Berry J A 1980 A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species. *Planta* 149, 78–90.
- Grand L F and Harvey A E 1982 Quantitative measurement of ectomycorrhizae on plant roots. In *Methods and Principles of Mycorrhizal Research*. Ed. N C Schenck. pp 157–164. American Phytopathological Society, St. Paul, MN.
- Graumlich L J 1991 Subalpine tree growth, climate, and increasing CO₂: An assessment of recent growth trends. *Ecology* 72, 1–11.
- Hollinger D Y 1987 Gas exchange and dry matter allocation responses to elevation of atmospheric CO₂ concentration in seedlings of three tree species. *Tree Physiol.* 3, 193–202.
- Hsiao T C, Acevedo E, Fereres E and Henderson D W 1976 Stress metabolism: Water stress, growth, and osmotic adjustment. *Phil. Trans. R. Soc. Lond. B* 273, 479–500.
- Kaushal P, Guehl J M and Aussenac G 1989 Differential growth response to atmospheric carbon dioxide enrichment in seedlings of *Cedrus atlantica* and *Pinus nigra* ssp. *Laricio* var. *Corsicana*. *Can. J. For. Res.* 19, 1351–1358.
- Kienast F and Luxmoore R J 1988 Tree-ring analysis and conifer growth responses to increased atmospheric CO₂ levels. *Oecologia* 76, 487–495.
- LaMarche Jr V C, Graybill D A, Fritts H C and Rose M R 1984 Increasing atmospheric carbon dioxide: Tree ring evidence for growth enhancement in natural vegetation. *Science* 225, 1019–1021.
- Lewis J D, Thomas R B and Strain B R 1994 Effect of elevated CO₂ on mycorrhizal colonization of loblolly pine (*Pinus taeda* L.) seedlings. *Plant Soil* 165, 81–88.
- Lincoff G H 1981 *The Audubon Society Field Guide to North American Mushrooms*. Alfred A. Knopf, New York. 926 p.
- Molina R and Trappe J M 1982 Patterns of ectomycorrhizal host specificity and potential among Pacific Northwest conifers and fungi. *For. Sci.* 28, 423–458.
- Norby R J 1989 Direct responses of forest trees to rising atmospheric carbon dioxide. In *Air Pollution Effects on Vegetation Including Forest Ecosystems: Proceedings of the Second US-USSR Symposium*, September 13–25, 1988, Corvallis, OR, Raleigh, NC and Gatlinburg, TN. Eds. R D Noble, J L Martin and K F Jensen. pp 243–249. USDA Forest Service Northeastern Forest Experiment Station, Broomall, PA.
- Norby R J 1994 Issues and perspectives for investigating root responses to elevated atmospheric carbon dioxide. *Plant Soil* 165, 9–20.
- Norby R J and O'Neill E G 1989 Growth dynamics and water use of seedlings of *Quercus alba* L. in CO₂-enriched atmospheres. *New Phytol.* 111, 491–500.
- Norby R J, O'Neill E G, Hood W G and Luxmoore R J 1987 Carbon allocation, root exudation and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO₂ enrichment. *Tree Physiol.* 3, 203–210.
- Norby R J, O'Neill E G and Luxmoore R J 1986 Effects of atmospheric CO₂ enrichment on the growth and mineral nutrition of

- Quercus alba* seedlings in nutrient-poor soil. *Plant Physiol.* 82, 83–89.
- Norby R J, Wullschlegel S D and Gunderson C A 1996 Tree responses to elevated CO₂ and implications for forests. In *Carbon Dioxide and Terrestrial Ecosystems*. Eds. G W Koch and H A Mooney. pp 1–21. Academic Press, New York.
- O'Neill E G, Luxmoore R J and Norby R J 1987a Elevated atmospheric CO₂ effects on seedling growth, nutrient uptake, and rhizosphere bacterial populations of *Liriodendron tulipifera* L. *Plant Soil* 104, 3–11.
- O'Neill E G, Luxmoore R J and Norby R J 1987b Increases in mycorrhizal colonization and seedling growth in *Pinus echinata* and *Quercus alba* in an enriched CO₂ atmosphere. *Can. J. For. Res.* 17, 878–883.
- Radoglou K M and Jarvis P G 1990 Effects of CO₂ enrichment on four poplar clones. I. Growth and leaf anatomy. *Ann. Bot.* 65, 617–626.
- Riffle J W 1973 Pure culture synthesis of ectomycorrhizae on *Pinus ponderosa* with species of *Amanita*, *Suillus*, and *Lactarius*. *For. Sci.* 19, 242–250.
- Rogers H H, Runion G B and Krupa S V 1994 Plant responses to atmospheric CO₂ enrichment with emphasis on roots and the rhizosphere. *Environ. Pollut.* 83, 155–189.
- Surano K A, Daley P F, Houpis J L J, Shinn J H, Helms J A, Palassou R J and Costella M P 1986 Growth and physiological responses of *Pinus ponderosa* Dougl. ex P. Laws. to long-term elevated CO₂ concentrations. *Tree Physiol.* 2, 243–259.
- Tissue D T, Thomas R B and Strain B R 1996 Growth and photosynthesis of loblolly pine (*Pinus taeda*) after exposure to elevated CO₂ for 19 months in the field. *Tree Physiol.* 16, 49–59.
- Tolley L C and Strain B R 1984 Effects of CO₂ enrichment and water stress on growth of *Liquidambar styraciflua* and *Pinus taeda* seedlings. *Can. J. Bot.* 62, 2135–2139.
- Walker R F, Geisinger D R, Johnson D W and Ball J T 1995a Interactive effects of atmospheric CO₂ enrichment and soil N on growth and ectomycorrhizal colonization of ponderosa pine seedlings. *For. Sci.* 41, 491–500.
- Walker R F, Geisinger D R, Johnson D W and Ball J T 1995b Enriched atmospheric CO₂ and soil P effects on growth and ectomycorrhizal colonization of juvenile ponderosa pine. *For. Ecol. Manage.* 78, 207–215.
- Waring R H and Cleary B D 1967 Plant moisture stress: Evaluation by pressure bomb. *Science* 155, 1248, 1253–1254.
- Yawney H W and Carl C M 1969 An improved seedling root-volume measuring device. *Tree Planters' Notes* 20, 7–8.

Section editor: E Garnier